

- PATENT COOPERATION EATY

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NOTIFICATION OF ELECTION

(PCT Rule 61.2)

Assistant Commissioner for Patents United States Patent and Trademark Office Box PCT Washington, D.C.20231

ÉTATS-UNIS D'AMÉRIQUE

Date of mailing (day/month/year) 09 August 1999 (09.08.99)	in its capacity as elected Office		
International application No. PCT/EP98/08522	Applicant's or agent's file reference DIR0550		
International filing date (day/month/year) 17 December 1998 (17.12.98)	Priority date (day/month/year) 24 December 1997 (24.12.97)		
Applicant BRANDS, Rudi			

1.	The designated Office is hereby notified of its election made:
	X in the demand filed with the International Preliminary Examining Authority on:
	17 June 1999 (17.06.99)
	in a notice effecting later election filed with the International Bureau on:
2.	The election X was
	was not
	made before the expiration of 19 months from the priority date or, where Rule 32 applies, within the time limit under Rule 32.2(b).

The International Bureau of WIPO 34, chemin des Colombettes 1211 Geneva 20, Switzerland

Authorized officer

Jean-Marie McAdams

Facsimile No.: (41-22) 740.14.35 Telephone No.: (41-22) 338.83.38

Form PCT/IB/331 (July 1992)

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(PCT Article 18 and Rules 43 and 44)

DIR0550 International application No. PCT/EP 98/ 08522	ACTION International filing date (day/month/year)	20) as well as, where applicable, item 5 below.
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	17/12/1998	24/12/1997
Applicant	,	
DUPHAR INTERNATIONAL RES	EARCH B.V. et al.	
This International Search Report has be	en prepared by this International Searching Auth	nority and is transmitted to the applicant
•	transmitted to the International Bureau.	
This International Search Report consis	ts of a total of <u>2</u> sheets. by a copy of each prior art document cited in this	report.
Basis of the report		
	e international search was carried out on the bas nless otherwise indicated under this item.	sis of the international application in the
the international search Authority (Rule 23.1(b))	was carried out on the basis of a translation of to.	he international application furnished to this
b. With regard to any nucleotide a was carried out on the basis of	and/or amino acid sequence disclosed in the in	ternational application, the international search
	tional application in written form.	
filed together with the ir	iternational application in computer readable form	n.
furnished subsequently	to this Authority in written form.	
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	ubsequently furnished written sequence listing das filed has been furnished.	oes not go beyond the disclosure in the
the statement that the infurnished	nformation recorded in computer readable form is	s identical to the written sequence listing has been
2. Certain claims were fo	ound unsearchable (See Box I).	
3. Unity of invention is la		
4. With regard to the title ,	•	
the text is approved as:	submitted by the applicant.	
the text has been estab	lished by this Authority to read as follows:	-
5. With regard to the abstract,		·
the text is approved as	submitted by the applicant.	
	lished, according to Rule 38.2(b), by this Authori he date of mailing of this international search rep	
6 The figure of the drawings to be pu	blished with the abstract is Figure No.	•
as suggested by the ap	plicant.	X None of the figures.
because the applicant f	ailed to suggest a figure.	
because this figure bett	er characterizes the invention.	



T/EP 98/08522

A. CLASS IPC 6	FICATION OF SUBJECT MATTER C12N5/00 C12N7/00		
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	o International Patent Classification (IPC) or to both national classific	cation and IPC	
Minimum de	ocumentation searched (classification system followed by classificat	ion symbols)	
IPC 6	C12N		
Documenta	tion searched other than minimum documentation to the extent that s	such documents are included in the fields se	earched
Electronic o	lata base consulted during the international search (name of data ba	ase and, where practical, search terms used)
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C. DOCUM	ENTS CONSIDERED TO BE RELEVANT		
Category °	Citation of document, with indication, where appropriate, of the re	levant passages	Relevant to claim No.
x	WO 92 10564 A (US ARMY ;CELLCO (US)).	1-6
	25 June 1992 see the whole document	·	
	see the whole document		
X	EP 0 417 531 A (BAYER AG) 20 Marc see the whole document	ch 1991	1-6
X	WO 89 08701 A (INST ANGEWANDTE		. 1–6
^	BIOTECHNOLOGIE) 21 September 1989	9	
	see the whole document		
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	her documents are listed in the continuation of box C.	V Patent family mambara are flated	in annay
		X Patent family members are listed	ın annex.
	stegories of cited documents:	"T" later document published after the inte or priority date and not in conflict with	the application but
consid	ent defining the general state of the art which is not dered to be of particular relevance document but published on or after the international	cited to understand the principle or the invention	
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other	ent referring to an oral disclosure, use, exhibition or means	document is combined with one or moments, such combination being obvious in the art.	
	ent published prior to the international filing date but han the priority date claimed	"&" document member of the same patent	family
Date of the	actual completion of the international search	Date of mailing of the international sea	arch report
2	7 April 1999	10/05/1999	
Name and	mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2	Authorized officer	
Ì	NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,	Hillenbrand, G	
	Fax: (+31-70) 340-3016	in inclinitation, d	

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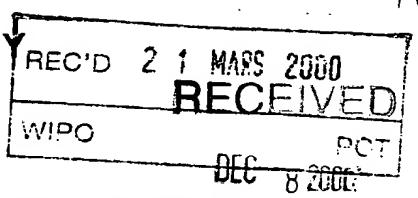
T/EP 98/08522

Patent document cited in search report		Publication date	Patent family member(s)		Publication date
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PATENT COOPERATION TREATY

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INTERNATIONAL PRELIMINARY EXAMINATION REPORT

TECH CENTO	1360,7509
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Applicants of OSSO	or agent's file reference		Notification of Transmittal of International Programme Programme (Form PCT/IPEA/416)
	I application No.	International filing date (day/month/year	Priority date (day/month/year)
PCT/EP9		17/12/1998	24/12/1997
nternationa C12N5/0	· ·	or national classification and IPC	
Applicant			
DUPHAF	INTERNATIONAL RES	SEARCH B.V. et al.	
		camination report has been prepared by tant according to Article 36.	his International Preliminary Examining Authorit
2. This F	REPORT consists of a total	al of 5 sheets, including this cover sheet.	
b	een amended and are the	anied by ANNEXES, i.e. sheets of the de basis for this report and/or sheets conta on 607 of the Administrative Instructions (scription, claims and/or drawings which have ining rectifications made before this Authority under the PCT).
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DEC 8 200C

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

TECH CENTURY (CENTURY INTERPORT No. PCT/EP98/08522

I.	Basis	of th	report
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1. This report has been drawn on the basis of (substitute sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to the report since they do not contain amendments.):

	•		lo not contain amendments.):
	Des	cription, pages:	
	1-10)	as originally filed
	Clai	ms, No.:	
	1-6		as originally filed
	Dra	wings, sheets:	
	1		as originally filed
2.	The	amendments hav	e resulted in the cancellation of:
		the description,	pages:
		the claims,	Nos.:
		the drawings,	sheets:
3.		This report has be considered to go	een established as if (some of) the amendments had not been made, since they have been beyond the disclosure as filed (Rule 70.2(c)):
1	Ada	litional observation	ns. if necessary:

- V. Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement
- 1. Statement

2. Citations and explanations

see separate sheet

VIII. Certain observations on the international application

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made:

see separate sheet

INTERNATIONAL PRELIMINARY **EXAMINATION REPORT - SEPARATE SHEET**

International application No. PCT/EP98/08522

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The examination is being carried out on the following application documents:

DEC 8 200C?

Text for the Contracting States:

AT BE CH DE DK ES FI FR GB GR IT IE LI LU MC NL PT SE

TECH CENTER 1600/2900

Description, pages:

1-10

as originally filed

Claims, No.:

1-6

as originally filed

Drawings, sheets:

as originally filed

Relevant documents cited

D1: WO 92 10564 A (US ARMY ; CELLCO (US)) 25 June 1992

Paragraph V (Reasoned statement):

Novelty (Article 33.2 PCT):

Having regard to the documents cited in the International Search Report the claimed matter is considered to be novel.

Inventive step (Article 33.3 PCT):

D1 describes the production of high titers of recombinant viral vectors and transduced target cells in a sustained and continuous process (see Fig. 2). The difference between D1 and the claimed matter appears to be the fact that the applicant claims a repeated discontinuous process. In view of D1 the IPEA considers carrying out the process described in D1 in a discontinuous manner by (a) using part of the cells of the preproduction batch for at least one production batch, and (b) using

INTERNATIONAL PRELIMINARY **EXAMINATION REPORT - SEPARATE SHEET**

the remaining part of the cells as a seed for the preparation of at least one subsequent preproduction batch as an obvious modification of the process described already in the cited prior art which does not require any inventive activity. This applies also to the use of a specific type of anchorage-dependent cells (see Claims 4-5). The fact that as a result of the claimed process it is possible to produce the product with cells at any passage number once a specific validation has been performed was also not surprising for the skilled person but the logical result of such a discontinues process.

Paragraph VIII (Certain observations on the international application):

The subject-matter of Claim 1 is too broadly and imprecisely drafted and thus does not comply with the requirements of Article 6 PCT. In Claims 4-5 the term "anchorage dependent" should be more clearly defined.

Finally, the attention of the applicant is directed to the fact that at present it appears that the claimed matter is not sufficiently clearly delimited from the teachings of US-A-5,017,490 and US-A- 4,664,912 (see page 1 of the description).

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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶:

C12N 5/00, 7/00

A1

(11) International Publication Number: WO 99/33955

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(21) International Application Number: PCT/EP98/08522

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97204110.7 24 December 1997 (24.12.97) EP

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(72) Inventor; and
 (75) Inventor/Applicant (for US only): BRANDS, Rudi [NL/NL];
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 36, NL-1381 CP Weesp (NL).

(74) Agent: BREEPOEL, Peter, Maria; Octrooibureau Zoan B.V., C. J. van Houtenlaan 36, NL-1381 CP Weesp (NL).

(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published

With international search report.

Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.

(54) Title: PREPARATION OF CELLS FOR PRODUCTION OF BIOLOGICALS

(57) Abstract

(30) Priority Data:

The present invention relates to a method for the preparation of cells for use in the production of biologicals, by culturing cells is a desired cell volume of a preproduction batch, where after in a repeated discontinuous process: a) part of the cells of the preproduction batch is used for the preparation of at least one production batch, and b) the remaining part of the cells of the preproduction batch is used as a seed for the preparation of at least one subsequent preproduction batch.

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WO 99/33955 PCT/EP98/08522.

Preparation of cells for pr ducti n of biologicals

The present invention is concerned with a method for the preparation of cells for use in the production of biologicals.

For the production of biologicals on e.g. cell lines, the preparation of large amounts of cells using an scaling up procedure in bioreactors will be necessary.

- The US patent No. 5,017,490 discloses such a scaling up procedure which provides in particular the advantage of a low risk of transfer contamination. This method is, however, not suited for anchorage dependent cells (hence, not for cells which only grow if fixed to a substrate) or cells embedded in a substrate (e.g. in porous carriers).
- The US patent No. 4,644,912 discloses a method for the preparation of anchorage-dependent cells for the production of biologicals (i.e. viruses) starting with a cell working seed, and with subsequent passages effected in increasing consecutive volumes of 1 litre, 5 litre, 25 litre, 150 litre bioreactors, and finally either in a 1000 litre bioreactor or in a multiplicity of 150 litre bioreactors. In between any of these passage steps the cells were released from their carriers with a dilute protease solution. In the final passage the inoculation by the virus was effected.

Assuming average cell cycle times of about 20-24 hours the passage intervals may be about every 3-5 day. Therefore, in order to expand the cells to sufficient large cultures from a MWCS¹ the total scaling up procedure may take several weeks, depending on the final bioreactor volume.

In the above methods for preparation of cells each of the ultimate production batches has to be prepared from the MWCS. For the production of vast amounts of biologicals it will be necessary to utilise several parallel culturing lines up to the largest vessel volumes. Such preparation procedure, hence, is very time consuming and necessitates the operation of a very considerable number of bioreactors for the preparation of the cells as well as for the production of the biologicals.

It is an object of the present invention to provide a much faster through-put in preparation of cells for the production of biologicals.

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¹ MWCS = manufacturer's working c !I bank

Accordingly, the present invention relates to a method for the preparation of cells for use in the production of biologicals, by culturing cells up till a desired cell volume of a preproduction batch, where after in a repeated discontinuous process:

a) part of the cells of the preproduction batch is used for the preparation of at least one production batch, and

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b) the remaining part of the cells of the preproduction batch is used as a seed for the preparation of at least one subsequent preproduction batch.

More in particular, the present invention relates to a method for the preparation of cells for use in the production of biologicals, by culturing cells up till a desired cell volume of a preproduction batch, where after in a repeated discontinuous process:

- a) part of the cells of the preproduction batch is transferred to be used for the preparation of at least one production batch, and
- b) the remaining part of the cells of the preproduction batch is transferred to be used as a seed for the preparation of at least one subsequent preproduction batch.

In a preferred embodiment of the present invention the first preproduction batch is prepared from a working seed stock by at least one passage step.

In a further preferred embodiment of the present invention the cells which are prepared are anchorage-dependent. In the latter case it will generally be necessary that the cells are grown on a substrate. It will then be advisable during the repeated process each time when part of a batch is used for the preparation of a new batch to add an additional amount of substrate. In a preferred embodiment, each time prior to the addition of substrate at least part of the cells are first released from their original substrate

As used herein the expression "production batch" means a culture of cells which is employed for the production of biologicals.

As used herein the expression "preproduction batch" means a culture of cells which is used in the process according to the present invention for the preparation of at least one production batch (as defined above) and one subsequent preproduction batch.

As used herein the expression "biological" means any substance or organism which can be produced from a cell culture. Examples of "biologicals" are viruses and proteins such as enzymes.

As used herein the expression "working seed stock" means an amount of a certain type of cells of defined ancestry stored to be used as a seed from which all cultures of the same type of cells are derived.

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As used herein the expression "anchorage-dependent cells" means cells which for their proper growing and/or propagation need to be attached to a substrate as defined herein.

As used herein the expression "substrate" means any particulate matter useful for the attachment of cells.

As used herein the expression "passage step" means a sequence of activities in the propagation and production of cells comprising at least the transfer of a suitable amount of cells and of a suitable amount of culturing medium into a production vessel, the incubation of the vessel at conditions suitable for the growing and propagation of the cells during a time sufficient for effective growing and propagation of the cells. Optionally a passage step may comprise separation of the cells from the culture medium and/or from the substrate after a time sufficient for effective growing and propagation of the cells.

It will be clear to the man skilled in the art that the method according to the present invention differs essentially from methods known in the art wherein cells are produced in a continuous process rather than the present discontinuous process. According to the patent publications EP0417531 and WO89/08701 continuous culture systems can be employed for the production of viruses as well. Firstly cells are grown in a first bioreactor, and after a certain cell density is reached cells are fed continuously from said first bioreactor into a second bioreactor. In this second bioreactor viruses are grown on the cells and subsequently these viruses are withdrawn continuously from this second bioreactor.

The basic method of working according to the present invention is to use a mother bioreactor from which the production bioreactor(s) is (are) fed with cells. When the cells are anchorage dependent, after each passage step cells preferably need to be detached from their substrates.

A trypsinisation procedure on large bioreactors has been developed for this purpose. The production cells are defined up to a specific and characterised passage number for a so-call d ECB². The method described allows high through-put production since the up scaling

² ECB = Extended Cell Bank

route from WCS to production cells can be very much shortened and much less bioreactors are needed since parallel production lines are not needed anymore.

Various embodiments of the present invention are depicted in Figure 1.

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In a preferred embodiment cells are expanded from one ampoule of a MWCS up to the level of the first preproduction batch through one or more passage steps. The size of the bioreactor used for such a preproduction batch can range from several litres working volume to several hundreds of litres. Next, a part e.g. 10-20 % of the cells thus expanded (e.g. passage X) are used to repopulate a bioreactor for the production of a subsequent preproduction batch (being passage number X+1), whereas the bulk of the cells is transferred (passage X or X+1) to a larger bioreactor size in order to start production directly or to first populate it, and subsequently start production.

In classical serial production lines the number of doubling of the cells derived from the MWCS at the moment of harvest is known up front within certain limits. A maximum allowable generation number is set to the production system at the onset.

In the method according to the present invention the maximum number of cell passages can be defined by ECB. Production passage number (the number of cell passages used prior to production of the biological product), hence, is irrelevant within the limits set by ECB. As a consequence, such maximum number of passages is to be obeyed in view of regulatory restrictions. As a result the particular batch of produces biologicals is the end product of one direct scaling up route.

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In order to verify whether the specifications of the cells at the stage of ECB in production are similar to the MCB³ one need to perform specific validation for this purpose with respect to growth characteristics, freedom of adventitious, extraneous and endogenous agents at the different stages, karyology iso-enzyme analysis and so on. Once such ECB is fully characterised one may allow to produce the product with cells at any passage number between MCB and ECB, since it may be assumed that cells have not changed in between in their specs. As a result tests on the MWCS therefore can be limited to sterility testing. This is a particular advantage of the method according to the present invention

³ MCB = Master Cell Bank

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With the maximum passage number set one may use cells at any stage in between. From this in order to further minimise the time needed to expand the cells from the MWCS to production bioreactor it would be an advantage to enable bulk start-up of cells. This can be done for example in one of the following ways:

• Cells may be parked at a certain passage number during longer intervals at ambient temperature (17-32 °C) and be revitalised to log expansion growth by raising the temperature and changing the culture medium, or

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• Cells may be frozen (Temp < -80°C) in bulk and be thawed prior to transfer them to a preset volume bioreactor, thereby reducing the needed up scaling route significantly.

The method according to the present invention can be carried out with animal cell cultures and more in particular with anchorage dependent cells. Suitable types of cells are e.g. hamster cells (CHO, BHK-1), monkey cells (Vero), bovine cells (MDBK), canine cells (MDCK), human cells (CaCo, A431) or chicken cells (CEF).

As a bioreactor according to the present inventions can be used a single unit of a plurality of units of e.g. stirred fermenters, fixed bed fermenters, fluidized bed fermenters, air lift fermenters, or a hollow fibre reactors.

- Cells of the above times can and some even should be cultured when fixed to a solid support, like micro-carriers or macro-carriers in suspension, e.g. in a fixed bed, a fluidized bed or in suspension, or like hollow fibres. Cells can also be embedded into a carrier (e.g. porous carrier)
- In the course of the method according to the present invention, in particular when using a solid support, cells are to be released from this solid support. This can be effected by any method useful for detaching of cells from a solid support. Advantageously, to this end use can be made of a proteolytic enzyme solution. Optionally, this enzymatic release step can be preceded by one or more pre-conditioning steps, e.g. by treatment with PBS and/or EDTA, in order to enhance the proteolytic efficiency, and/or in order to reduce the amount of proteolytic enzyme required.

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EXAMPLE 1

Cell detachm nt and separation from carriers prior to transf r to next bioreactor

- Anchorage dependent cells of a MDCK⁴ cell line were cultured at 37 °C on Cytodex-3 micro carriers (Pharmacia, Uppsala, Sweden) (5 g of carriers/I) in a stirred bioreactor of 4 litre ("mother bioreactor"). The growth medium was EpiSerf (Life Technologies, Paisly, Scotland). Growth was continued till a maximum of 5x10⁶ cells/ml of culture.
- The cells were detached from the carriers by trypsinisation in a Trypsin-EDTA solution (Life Technologies, Paisly, Scotland).
 - After settling of the carriers 80% of the detached cells were transferred to 3 other bioreactors of similar size. The latter "production" bioreactors all have carriers (cell substrate) added to them up front. Cells were allowed to repopulate the carriers and subsequently used for production in these production bioreactors.
- The remainder of the cells in the "mother bioreactor" were allowed to repopulate the remaining Cytodex-3 carriers and were cultured to the desired cell density.

EXAMPLE 2

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Cell detachment without separation from carriers prior to transfer to next bioreactor

The culturing of cells was carried out as described in Example 1, however after trypsinisation 80% of the detached cells including the carriers are transferred to the 3 production bioreactors. Additionally, suitable carriers were added to all bioreactors.

⁴ MDCK = Madin Darby Canine Kidney (cell line)

EXAMPLE 3

Cell detachm nt without separation from carriers after transfer to next bior actor

The culturing of cells was carried out as described in Example 1, however, 80 % of still adhered cells were transferred to a bioreactor of similar size which next was used directly for product generation.

The remaining cells on micro carriers in the mother fermenter were next detached by trypsinisation, where after new carriers were added and cells were allowed to repopulate the substrates.

EXAMPLE 4

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Start-up from frozen bulk cells

In this experiment part of the culture was used to rebatch the mother fermenter and some daughter fermenters and part of the culture was used to freeze cells in bulk.

Frozen bulk cells (total 14.4X10⁸ cells) were inoculated in a start culture in a 3 litre mother fermenter containing 5 g Cytodex per litre and EpiSerf medium, and thereafter incubated at 37 °C. Residual cryo-preservatives were removed by a medium change on day 1.

At day 2 trypsinisation was carried out, 50% of the cells were bulk frozen and the remaining cells were inoculated to micro-carriers in a subsequent fermenter.

From Table 1 it can be deduced that the cells do continue to grow at a normal rate between day 2 and 3

On day 4 the content of the mother fermenter was trypsin-detached and rebatched onto new micro-carriers (10 g/l) in two other fermenters next to the mother fermenter.

At day 5 the plating efficiency turned out to be about 85%.

Tabl 1

	3 litre mother ferm nt r	3 litre ferm nt r	3 litre ferment r
day	c ils x 100.000/mi	cells x 100.000/ml	cells x 100.000/ml
0	NOD		
1	6.6		
2	14		
3	15.5		
4	30		
5	5.5	.10	10
plating efficiency	85%	85%	85%

EXAMPLE 5

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Transfer from small scale mother fermenter to large scale production fermenter

Cells were scaled up to a large scale in 65 litre and 550 litre fermenters (50 litre and 250 litre working volume, respectively) using a micro-carrier density of 5 g Cytodex per litre.

As can be seen from Table 2, 90% of the total of cells is transferred to the large scale fermenter from a 50 litre fermenter culture with 800.000 cells/ml of which 69% proved to be viable.

The same was found in the 50 litre mother fermenter; about 69% of the repropagating cells turned out to be viable.

15 The procedure was as follows:

On day 0, the carriers were allowed to settle in the 50 litre culture, where after the supernatant (culture medium) was removed and replaced by PBS. The content of the fermenter was agitated for 5-15 minutes. The supernatant was removed after resettling of the carriers. This step can be repeated if needed.

Next this step was repeated with PBS/EDTA (0.4 gram EDTA/litre PBS). Again the culture was agitated during 5-15 minutes, carriers were allowed to settle, the supernatant was removed, and the PBS/EDTA step was repeated until cells had become rounded and were ready to be trypsin-detached.

Then trypsin (0.025% final concentration) was added to the PBS/EDTA and incubated for 5-15 minutes. Next either the cell containing supernatant (after settling of now "nude" carriers) were transferred (as in xample 9) or the mixture of cells plus carriers were transferred (total 80 % of total mix).

After transfer of the cells to the 550 litre fermenter the remainder of the cells (hence, 10% of the viable cells) were allowed to repopulate the carriers still present in the fermenter after refilling the 50 I fermenter with culture medium.

About 70% of the cells proved to be viable

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Table 2

	50 litre culture	250 litre culture
day	cells x 100.000/ml	cells x 100.000/ml
0	8 (400 x 10 ⁸ total cells)	1.1 (275 x 10 ⁸ viable cells)
1		0.8
2		2.9
3		3.4
4		8.9
5		18.0

EXAMPLE 6

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Analogous to Example 5, however, 80% of the culture of the carrier-bound cells were transferred from the mother bioreactor to the production bioreactor. Production was started after addition of virus.

The 20% of cells and carriers remaining in the mother bioreactor were trypsinized and detached and upon addition of new substrate into the mother bioreactor were allowed to repopulate the mother bioreactor while production is ongoing in the physically separated production bioreactor.

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EXAMPLE 7

Large scale culture started from bulk frozen cells.

Bulk frozen cells were thawed and inoculated on a 10 litre (working volume) fermenter (Cytodex carrier density 5 g/l; culture medium EpiSerf) at an inoculation density of 1x10⁶ cells/ml. After attachment, the culture medium was replaced in order to remove residual cryoprotectants.

After day 1 the amount of viable cells attached to the carriers was 0.45x10⁶ cells/ml which from then on started growth. At a density of 2.8x10⁶ cells/ml the cells were detached from their

carriers by trypsinisation and 80 % was transferred to a 50 litre working volume fermenter (carriers 5 g/l).

As can be deduced from Table 3, at day 1 the amount of viable cells after bulk freezing of cells was about 45 %.

5 Of the total amount of transferred cells, the viability after trypsin detachment was 71.4%.

Table 3

day	cell density (x 10 ⁶ /l) in:			
	10 litre fermenter	50 litre fermenter		
0	1.0			
1/2	0.45			
3/4	1.3			
5	2.6			
6	2.8 $(280 \times 10^8 \text{ total})$			
6	0.6 (60 x 10 ⁸ total)	0.28 (140 x 10 ⁸ total)		
7		0.4 (200 x 10 ⁸ total)		

Claims

1. Method for the preparation of cells for use in the production of biologicals, by culturing cells up till a desired cell volume of a preproduction batch, where after in a repeated discontinuous process:

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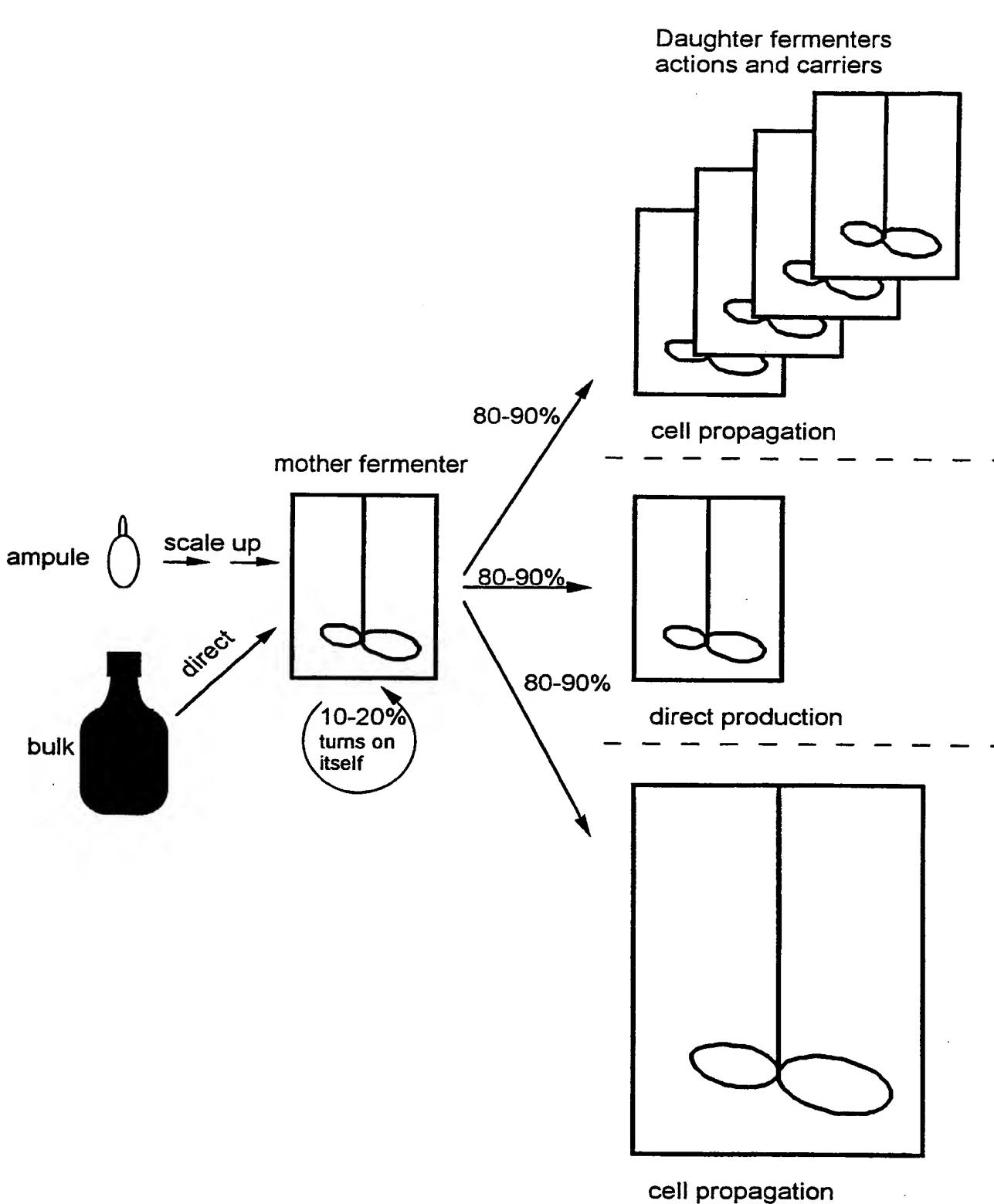
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- a) part of the cells of the preproduction batch is used for the preparation of at least one production batch, and
- b) the remaining part of the cells of the preproduction batch is used as a seed for the preparation of at least one subsequent preproduction batch.
- 2. Method according to claim 1 wherein in the repeated discontinuous process:
 - a) part of the cells of the preproduction batch is transferred to be used for the preparation of at least one production batch, and
 - b) the remaining part of the cells of the preproduction batch is transferred to be used as a seed for the preparation of at least one subsequent preproduction batch.
- Method according to claim 1 or 2, characterised in that a first preproduction batch is prepared from a working seed stock by at least one passage step.
 - 4. Method according to claim 1-3, characterised in that the cells are anchoragedependent.
- Method according to claim 2, characterised in that the cells are anchorage dependent, the cells are grown on a substrate, and prior to each transfer step the cells are released from their substrate.
 - 6. Method according to claim 1-5, characterised in that the biological of interest is a virus.

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FIGURE 1



cell propagation add new carriers

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A. CLASS IPC 6	C12N5/00 C12N7/00			
According	to International Patent Classification (IPC) or to both national cla	ssification and IPC		
B. FIELDS	S SEARCHED			
Minimum d IPC 6	documentation searched (classification system followed by class ${\tt C12N}$	ification symbols)		
Documenta	ation searched other than minimum documentation to the extent	that such documents are included in the fields s	earched	
Electronic	data base consulted during the international search (name of da	ta base and, where practical, search terms used	1)	
C DOCUM	AENTS CONSIDERED TO BE BELLEVIANT	-		
Category °	MENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document, with indication, where appropriate, of the	e relevant passages	Relevant to claim No.	
X	WO 92 10564 A (US ARMY ;CELLCO 25 June 1992 see the whole document	(US))	1-6	
X	EP 0 417 531 A (BAYER AG) 20 M see the whole document	arch 1991	1-6	
X	WO 89 08701 A (INST ANGEWANDTE BIOTECHNOLOGIE) 21 September 1989 see the whole document		1-6	
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	her documents are listed in the continuation of box C.	χ Patent family members are listed	in annex.	
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Date of the	actual completion of the international search	Date of mailing of the international sea	arch report	
2	7 April 1999	10/05/1999		
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